

TWO

The Biology of Successful Aging: Watchful Progress at Biogerontology's Known–Unknown Interface

David J. Waters and Naomi N. Kariuki

This chapter is about the biology of aging. It offers a contemporary perspective on the kinds of ideas that biogerontologists wonder about. As investigators, we are particularly keen to highlight individual lines of inquiry that are leading to discoveries about the aging process. We know of no better way of realizing this goal than to situate our subject—the biological underpinnings of healthy longevity—as a moving target, a field in flux. Accordingly, this chapter sets out to sketch in sufficient detail the rich evolution of ideas in this ever-moving field of aging research, underscoring substantial advances and critical gaps in our understanding.

In this chapter, the term *organismal senescence* is used to describe the deteriorative changes that result in decreased viability and ultimately an increase in an organism's risk for mortality. In contrast, the term *aging* refers to any time-dependent changes that occur during the life course of an organism. Thus, aging encompasses changes that are good (e.g., increased wisdom), bad (e.g., arteriosclerosis), or inconsequential (e.g., baldness) in terms of their effect on viability and survival (Finch, 1990). It is an unfortunate reality that scientists and the public often use these terms interchangeably, resorting to using the term *aging* synonymously with deterioration.

Not all organisms undergo senescence at the same tempo (Finch, 1990). Humans exhibit gradual senescence (deterioration over decades), whereas other species, such as bamboo, have very rapid senescence (deterioration over days). There is an eclectic group of

organisms that exhibit negligible senescence; age-related deteriorative changes are virtually imperceptible in these species. Not surprisingly, organisms with negligible senescence—such as rockfish, bristlecone pine, and possibly the naked mole rat—are currently the subject of great scientific interest (Finch, 1998, 2009; Finch & Austad, 2001; Pérez et al., 2009).

Understanding the mechanisms of organismal senescence and how morbidity can be compressed, culminating in extension of a healthy life span, is the goal embraced by most biogerontologists (Fries, 2003; Olshansky, Hayflick, & Carnes, 2002). These aims must be clearly articulated to the public (Binstock, 2004). It is also critical that the public understands what the goals of biogerontology research are not. For example, reversal of aging and achieving immortality are not considered tenable objectives. Including these metrics in the scientific dialogue serves only to compromise the public's understanding of the purpose and value of biogerontology research.

Basic and applied scientists are working diligently to understand the processes of cellular and organismal senescence and to identify the determinants of successful human aging. Some of the domains attracting the deepest inquiry, along with instructive examples of the diverse and complementary scientific approaches at hand, are framed in the sections that follow.

LESSONS FROM EVOLUTIONARY BIOLOGY

It has been argued that longevity, not aging or senescence, per se, is under genetic control (Martin, Austad, & Johnson, 1996). These so-called longevity genes extend longevity by increasing physiological reserve or increasing disease resistance rather than having a direct effect on the intrinsic rate of aging. Evolutionary biologists would concur that genes that regulate late-life deterioration would not be under the influence of natural selection. Instead, natural selection favors genes that influence the likelihood of reproductive success, because organisms rely on the successful completion of development and reproduction to perpetuate the species. During the postreproductive period, however, the force of natural selection quickly diminishes, rendering organisms ill-equipped to protect themselves against the age-related accumulation of molecular damage. Seldom is this an important consequence to animals living in the wild because, in general, they experience relatively brief postreproductive life spans, owing to their vulnerability to predators, accidents, or infectious diseases. In contrast, humans and other highly protected, domesticated animals encounter the manifestations of organismal senescence, including cancer and other age-related degenerative diseases.

When it comes to age-related deterioration and disability, two tenets of evolutionary theory would suggest that humans and other domesticated species are indeed hardwired for postreproductive calamity. First, the *disposal soma hypothesis* (Kirkwood, 1977, 1990) states that after reproduction, the soma (body) can be thrown away. Second, genes that confer an advantage early in life may also exert detrimental effects in the postreproductive period—the theory of *antagonistic pleiotropy* (Williams, 1957). The incidence of several age-related diseases is likely to be under the control of such antagonistically pleiotropic genes. For example, genes that stimulate abundant blood vessel formation within the placenta—favoring successful reproduction early in life—might be considered detrimental later in life if they promote a rich supply of blood vessels to rapidly growing tumors. It is expected that the frequency of alleles that exert these late-life detrimental effects would be maintained or even increased in the population owing to their early-life benefit.

Thus, organismal senescence and accompanying age-related diseases are an inevitable by-product of domestication (see Figure 2.1). An organism is not built with any regard for optimizing aging. Instead, an organism is designed to successfully and advantageously complete its maturation and reproduction. As a consequence, in species attaining a prolonged postreproductive life span, one expects to see an accumulation of molecular disorder (Hayflick, 1998). One such contributor to this molecular disorder is the accumulation of misfolded proteins resulting from the

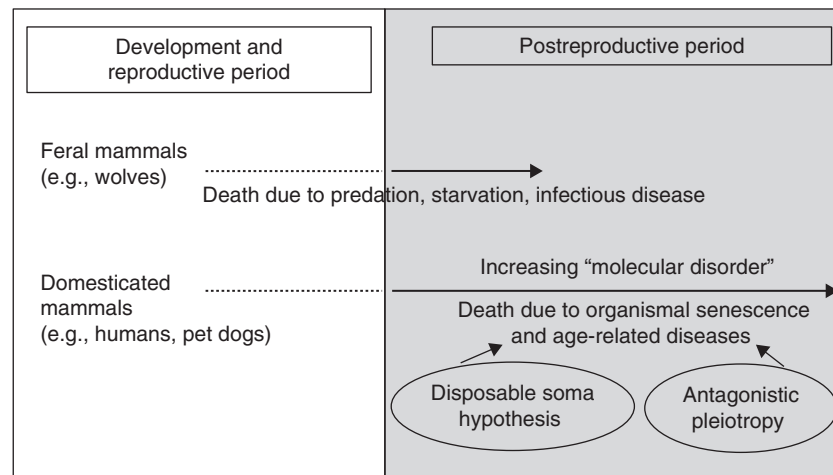


FIGURE 2.1 Domestication's curse: organismal senescence and the incidence of age-related diseases.

age-associated erosion in protein quality control—termed *proteotoxicity* (Koga, Kaushik, & Cuervo, 2011). Mechanistically, both dysregulation at the level of the proteasome and autophagy have been implicated in this disordered physiology (Cuervo, 2010; Powers, Morimoto, Dillin, Kelly, & Balch, 2009; Wong & Cuervo, 2010).

LESSONS FROM THE CELL CULTURE LABORATORY

Fifty years ago, Hayflick and Moorhead (1961) advanced the notion that normal somatic cells have a finite replicative life span in cell culture. Before this, it was believed that normal cells in culture, if cared for properly, could divide indefinitely. Subsequent studies have provided solid support for the concept of *replicative senescence*, whereby normal diploid cells have a limited number of population doublings (known as the Hayflick limit).

The process of in vitro replicative senescence has been studied extensively in normal fibroblasts and other cell types in cell culture (Campisi, 1996; Cristofalo & Pignolo, 1993). Not only do senescent cells suffer irreversible loss of their ability to replicate, they undergo other phenotypic changes. For example, senescent cells increase their production of matrix metalloproteinase enzymes, which might be expected to alter the function of tissues by favoring the turnover of extracellular matrix and tissue remodeling. It has been proposed that the senescence-associated secretory phenotype (SASP) may promote age-related pathologies, such as cancer and inflammation (Davalos, Coppe, Campisi, & Desprez, 2010; Rodier & Campisi, 2011). Importantly, *senescent cells do not die*; in fact, they retain their viability and are actually resistant to apoptotic cell death.

The in vitro replicative life span of normal cells is based on the number of population doublings rather than elapsed time in culture. The number of potential population doublings that a cell can undergo is determined by the length of *telomeres*, the structures that cap the ends of chromosomes (Kim, Kaminker, & Campisi, 2002). When normal cells undergo cell division, each replication witnesses a shortening of the telomere. When telomeric shortening reaches a critical threshold, the ability to replicate is lost; this signal is presumably mediated by a cellular DNA damage response (Dimri, 2005). In contrast, stem cells and most tumor cells possess the enzyme telomerase, which effectively maintains telomere length and enables these cells to escape the rules of replicative senescence (Granger, Wright, & Shay, 2002). Because cellular senescence is linked to telomere shortening, telomere length is actively being investigated as a biomarker to predict successful human aging (Bakaysa et al., 2007; Epel et al., 2004; Mather, Jorm, Parslow, & Christensen, 2011; Song et al., 2010).

There are at least two lines of evidence that make it attractive to consider that aging in the cell culture laboratory is indeed relevant to organismal senescence. First, when one looks across different species, the population doubling potential for cultured normal fibroblasts is proportional to the maximum life span potential of animals (Hayflick, 1976). Second, in studies with human fibroblasts, there is an inverse correlation between the age of the donor and replicative life span of cultured cells (Martin, Sprague, & Epstein, 1970). However, a study by Cristofalo and colleagues (Cristofalo, Allen, Pignolo, Martin, & Beck, 1998) challenged this notion by showing no significant relationship between donor age and the replicative potential of human fibroblasts. Moreover, the persistence of high replicative capacity in skin fibroblasts from 90-year-old people casts further doubt on the inverse association between donor age and growth characteristics of cultured fibroblasts (Maier et al., 2007). One possible explanation for these contradictory results is that *in vivo* aging does not result in a global, uniform, age-related loss of proliferative capacity. Instead, *in vivo* aging might be characterized by *mosaicism*—with old tissues representing a mosaic mixture of senescent and “youthful” cells. If true, then even old hosts have populations of youthful fibroblasts with a replicative life span comparable to that of cells retrieved from young hosts.

However, it is still not clear whether senescent cells actually accumulate *in vivo* in older individuals. Using the enzyme senescence-associated beta-galactosidase (Sa β gal) as a marker of cellular senescence, Dimri and colleagues (1995) found more senescent cells in the skin of elderly people than in younger subjects. Although the species and tissue specificity of Sa β gal activity are yet to be fully characterized, work with Sa β gal is addressing the critical need for biomarkers of senescence so that we can determine the extent to which the accumulation of senescent cells causes the functional decline seen in old tissues and old organs (Debacq-Chainiaux, Erusalimsky, Campisi, & Toussaint, 2009; Lee et al., 2006).

Now, biogerontologists working in the cell culture laboratory are moving beyond replicative senescence, extending their research techniques in important ways. They are beginning to carefully catalog the resistance of cells to *ex vivo* challenge with lethal and sublethal stresses as a new approach to getting a cellular handle on what it takes to achieve extended longevity. Testing the resilience of skin fibroblasts cultured from an array of beasts and bipeds is yielding a collection of provocative, insult-specific responses that, once deciphered, will undoubtedly point to new leads, fresh hypotheses to pursue (Dekker et al., 2009; Finch, Morgan, Longo, & de Magalhaes, 2010; Harper, Salmon, Leiser, Galecki, & Miller, 2007; Miller, Williams, Kiklevich, Austad, & Harper, 2011).

DETERMINANTS OF LIFE SPAN: GENES, ENVIRONMENT, AND CHANCE

Evolutionary theory teaches us that there are no genes whose purpose is to cause aging or accelerate the rate of aging. In that sense, senescence is not programmed, but rather a passive by-product of an organism's genetic hardwiring. That genes influence life span is not subject to debate—no amount of environmental protection or enrichment can produce a rat that lives as long as the average human. Specific genes have been identified that regulate life span in worms (*Caenorhabditis elegans*) (Kimura, Tissenbaum, Liu, & Ruvkun, 1997; Morris, Tissenbaum, & Ruvkun, 1996), flies (*Drosophila*) (Clancy et al., 2001; Lin, Seroude, & Benzer, 1998), and mice (Migliaccio et al., 1999). What remains enigmatic is whether any of these genes actually prolongs life span by slowing down the rate of aging.

Data from twin studies (Ljungquist, Berg, Lanke, McClearn, & Pedersen, 1998; McGue, Vaupel, Holm, & Harvald, 1993) suggest that 25% to 30% of the variation in life span among individuals can be attributed to genetic factors. In other words, *there is considerable plasticity to life span*. This means lifestyle is important—both the external environment (e.g., diet, exposure to chemicals) and the internal environment (e.g., hormones, oxidative stress) significantly influence an individual's life span.

Recently, the widely accepted idea that gene–environment interactions are the major determinant of all complex traits, including longevity, has made room for a third element: chance. A complex choreography of genes, environment, and chance orchestrates a vast array of biological outcomes (reviewed in Finch & Kirkwood, 2000). That chance has an important impact on life span is supported by observations that genetically identical organisms raised under identical conditions have differences in life span—all of the individuals do not drop dead on the same day. An elegant study in worms underscores the impact of intrinsic chance in determining longevity (Rea, Wu, Cypser, Vaupel, & Johnson, 2005).

Although several organismal models suggest that life span is indeed under genetic control, there are differing views about how many genes regulate longevity and how those genes should be classified. Martin (1978) estimates as many as 7,000 genes may regulate life span, suggesting that longevity is under polygenic control. However, some scientists favor the idea of oligogenic control—a few critical genes may account for the lion's share of the 25% to 30% portion of human life span that is heritable. This view is supported by a considerable body of evidence from *C. elegans* in which single gene mutations can significantly prolong life span (Kimura et al., 1997; Morris et al., 1996).

Today, the search continues for genes that can persist within natural populations and increase life span by decreasing the rate of aging. Although unproven, the existence of such genes is supported by the profound within-species difference in life span observed in island- versus mainland-dwelling opossums (Austad, 1993), selective breeding experiments in flies (Arking, 1987), and the natural evolution of dog breeds (Deeb & Wolf, 1994; Patronek, Waters, & Glickman, 1997; Waters & Wildasin, 2006). These so-called longevity assurance genes (Butler et al., 2003; Hodes, McCormick, & Pruzan, 1996) are assumed to influence the rate of aging because they are beneficial to the organism, for example, by regulating antioxidant defenses or DNA repair. However, scientists in hot pursuit of candidate longevity assurance genes are bound to uncover some unexpected results, because these genes will likely have tissue-specific effects that manifest themselves differently during different “windows” of the life course. Furthermore, nonlinear U-shaped dose responses and threshold effects, in which further increases in gene action do not enhance the organism’s viability, may befuddle investigators (Chiang et al., 2010; Kizer et al., 2011; Munhoz, Sorrells, Caso, Scavone, & Sapolsky, 2010; Wu, Cypser, Yashin, & Johnson, 2008).

To better understand the genetic basis of health, investigators are now looking beyond genetic variants to the impact of *epigenetic changes*. Epigenetic changes are transmissible alterations in gene expression caused by mechanisms other than variations in DNA sequence (Aguilera, Fernández, Muñoz, & Fraga, 2010). Differences in epigenetic programming—the capacity of an organism to turn on or off genes in response to environmental signals such as maternal care—provide a mechanistic basis to explain, in part, striking interindividual differences in resilience and healthy longevity (Weaver et al., 2004). Discovering which aging-related processes are regulated by epigenetic mechanisms is considered a high priority, because each individual’s epigenetic state is potentially reversible—not hardwired like genomic DNA sequences (Gravina & Vijg, 2010). Results from rodent studies showing the ability to experimentally manipulate epigenetic changes—using drugs in adulthood to reverse adverse stress behaviors caused by low-quality maternal care earlier in life—open the door to almost limitless possibilities to investigators thinking epigenetically (Weaver, Meaney, & Szyf, 2006).

HORMONES AND AGING: ALMOST NOTHING IS RAGING

The endocrine system, a major contributor to the host’s internal environment, is profoundly and predictably affected by aging. An age-related decline in hormone production results in lower circulating

levels of growth hormone/insulin-like growth factor-1 (IGF-1) (somatopause), estrogen (menopause), testosterone (andropause), and DHEA (adrenopause) (Lamberts, van den Beld, & van der Lely, 1997). The age-associated decline in these regulators of homeostasis has several important biological and clinical implications.

Although unproven, the idea that alterations in hormone levels cause organismal senescence, including the physiological deterioration seen in older adults, is supported by clinical observations. For example, loss of estrogen action in postmenopausal women has been associated with an increased risk for cardiovascular disease, accelerated loss of bone mass, and a decline in cognitive function (Love et al., 1992). In addition, individuals suffering from inherited growth hormone deficiency show many signs that mimic normal aging, such as thinning skin and decreased bone and muscle mass (Rudman et al., 1990). It follows from these observations that hormone supplementation might prove a useful intervention to significantly decrease the rate of age-associated physiological decline.

From a biological standpoint, there are several issues that transform successful hormonal replacement from a straightforward proposition to a formidable task. First, one must decide whether to use a *replacement* dose (i.e., restoring circulating hormone concentrations in older adults to normal youthful levels) or *supraphysiologic* dose (i.e., achieving high-circulating hormone concentrations that exceed normal levels). Other issues, such as timing of hormone administration during the life course (Bartke et al., 1998), use of a single hormone or combination (Shumaker et al., 2003; Tang et al., 1996), and sex-specific effects (Blackman et al., 2002) add another layer of complexity.

Second, hormones have *tissue-specific actions*—some desirable, some undesirable. For example, testosterone supplementation may decrease frailty in older men by increasing muscle mass and muscle strength (Krause, Mueller, & Mazur, 2005). However, an expected downside of testosterone supplementation is an increased incidence of prostatic disease. Because of potentially troubling trade-offs, scientists are developing hormonal agents with tissue-selective actions. For example, selective estrogen receptor modulators (SERMs), such as tamoxifen and raloxifene, exert beneficial effects on particular target organs (e.g., brain, blood vessels, bone) without detrimental stimulatory effects on cancer cells in the breast or uterus (Jordan, 2004).

Finally, it will be important to develop reliable biomarkers that can identify those individuals who will benefit most from hormone supplementation. By accomplishing this, it will be possible to finger hormone supplementation regimens that have the best risk–benefit profile (Bhasin et al., 2010).

It is not clear, however, whether an age-related decline in circulating hormones is in actuality a bad thing. Take growth hormone for example. If decreased growth hormone production contributes significantly to the age-related decline in muscle mass and bone density, then a decline in circulating growth hormone levels would be seen as maladaptive. On the other hand, the age-associated drop in growth hormone might serve as a favorable anticancer mechanism. Decreased growth hormone translates into lower levels of circulating IGF-1, a peptide that stimulates the survival and proliferation of most human breast, prostate, and colon cancers (Chan et al., 1998; Giovannucci et al., 2000; Schernhammer, Holly, Pollak, & Hankinson, 2005). Experimentally, carcinogen-treated mice with growth hormone deficiency are cancer resistant; growth hormone infusions render these mice more vulnerable to cancer development (Ramsey et al., 2002). Studies across several species have linked *low somatotropic tone*—a relative reduction in GH and IGF-1 action—with increased longevity (Bartke, 2008; Piper, Selman, McElwee, & Partridge, 2008). The association between exceptional human longevity and loss-of-function mutations in the IGF receptor, leading to diminished IGF-1 signaling, is consistent with this notion (Suh et al., 2008).

CALORIC RESTRICTION: IMPRACTICAL INTERVENTION, INVALUABLE RESEARCH TOOL

In 1935, McCay and colleagues reported that restricting food intake delayed the onset of age-related diseases and extended the life span of rodents (McCay, Crowell, & Maynard, 1935). Seventy-five years later, caloric restriction (CR, also called undernutrition without malnutrition or dietary restriction) is regarded as the most robust experimental intervention for extending life span in rodents (Masoro, 2003). The beneficial effects of CR on life span extension and cancer suppression are observed when total energy intake is curtailed to 20% to 40% less than the caloric intake of ad libitum fed animals. The experimental paradigm of CR is distinct from starvation: CR animals receive fewer calories but nutritionally adequate levels of all essential nutrients.

Most experiments in rodents employ lifelong CR initiated soon after weaning. However, life span extension in mice has been documented when CR was initiated at middle age, that is, 12 months old (Pugh, Oberley, & Weindruch, 1999). Both CR rodents and CR monkeys show significant improvements in several physiological parameters, including decreased fasting blood glucose and insulin levels (Roberts et al., 2001). Recent results showing that CR reduces age-related mortality

and disease incidence in nonhuman primates have fueled further inquiry (Colman et al., 2009).

However, a study of 41 different recombinant inbred strains of mice has refocused attention on a glaring deficiency in our understanding of just who will benefit from CR (Liao, Rikke, Johnson, Diaz, & Nelson, 2010). The surprising result: *The majority of mouse strains showed no life extension with CR*. In fact, CR *shortened* the life span of more strains than it lengthened. It is safe to say that the previously held notion that life span extension by CR is a universal outcome has been thrown under the bus (Harper, Leathers, & Austad, 2006; Liao et al., 2010). Now investigators are turning these genetically heterogeneous mouse strains into powerful tools of discovery—deciphering which genes are linked to differences in the longevity-extending effects of CR (Liao et al., 2011).

The exact mechanism by which CR extends life span is under intense investigation. Newer research is even beginning to explore the radical notion that CR is a form of dietary imbalance whose effects may be more closely linked to amino acid balance than calories (Grandison, Piper, & Partridge, 2009; Piper, Partridge, Raubenheimer, & Simpson, 2011). By identifying key targets modulated by CR, scientists hope to identify critical regulatory points that control senescence and longevity. One thing is certain: CR exerts pleiotropic effects, significantly altering numerous biochemical, immunologic, and hormonal networks.

The dogged search for the mechanistic basis for the longevity-enhancing effects of CR is more than just an academic exercise. Scientists are avidly seeking to develop CR mimetics because, long-term, 20% to 40% CR is an impractical intervention strategy to implement in humans (Hass et al., 1996). The discovery of effective CR mimetics would allow individuals to reap the benefits of CR without the undesirable effects of severe food restriction. However, because of our rudimentary understanding of how CR really works, there is considerable scientific debate as to what kind of agents would be the most effective CR mimetics.

Finally, as an alternative to sustained CR, *intermittent* CR might provide a more practical lifestyle modification. Currently, it is unclear whether intermittent CR would effectively mimic the beneficial effects of CR. Several lines of evidence indicate that alternate day fasting has different effects on both hippocampal and gonadal gene expression compared with an isocaloric CR regimen enforced every day (Martin et al., 2008; Martin et al., 2009). Alternate-day fasting exerts beneficial effects on blood glucose regulation and insulin sensitivity in mice without the reduction in body weight that typically accompanies CR (Anson et al., 2003). Moreover, intermittent fasting has been

linked to cardioprotection and cancer preventive effects in rodents (Varady & Hellerstein, 2007). In the future, we can expect further studies designed to establish the potential for intermittent CR to reduce disease risk and promote healthy longevity in humans.

RUSTING OUT: THE OXIDATIVE STRESS HYPOTHESIS

In 1956, Denham Harman proposed that endogenous products of aerobic metabolism are a major determinant of organismal senescence (Harman, 1956). Harman's free radical theory of aging predicted that organisms would inevitably suffer age-related accumulation of macromolecular damage induced by reactive oxygen species (ROS), leading to physiological decline and increased risk of mortality. Over the ensuing half century, substantial correlative data have been collected from animal studies that support this popular hypothesis, although definitive evidence that oxidant stress causes organismal senescence is lacking. Elucidating the precise role of oxidative stress in the aging process is complicated by the fact that the impact of oxidative stress on an organism's well-being depends on: (a) the amount of ROS production; (b) the vulnerability of macromolecules to ROS-induced damage, which reflects cellular antioxidant defenses; and (c) the capacity to repair oxidative damage (Beckman & Ames, 1998). Moreover, emerging data suggest that conceptualizing oxidative stress as "good" or "bad" is an ill-advised oversimplification—one that could ultimately mislead our assumptions about oxidative stress and aging.

Sources of Reactive Oxygen Species

The mitochondria represent the most important cellular source of ROS. The electron transport chain within mitochondria generates the adenosine triphosphate (ATP) necessary to power the cell, but is an imperfect system, resulting in the generation of ROS, especially hydroxyl radical. Oxidative damage to mitochondrial DNA leads to the production of defective mitochondrial proteins, which favors impaired electron transfer and a resultant increase in ROS production. According to the mitochondrial theory of aging (Miquel, 1992), the age-related accumulation of mitochondrial DNA mutations contributes significantly to organismal senescence by driving a vicious cycle of mitochondrial dysfunction and cellular energetic collapse. There are also important non-mitochondrial sources of oxidants, including beta oxidation of fatty acids, cytochrome p450 enzyme, and the oxidative burst used by phagocytic cells to destroy pathogens.

Macromolecular Targets and Measures of Oxidative Stress

Lipids, proteins, and nucleic acids are vulnerable to damage by oxidative stress. Lipid peroxidation leads to changes in membrane fluidity that can corrupt the cell's signal transduction—that is, how a cell processes information from its environment and interacts with other cells (Monteiro & Stern, 1996). The level of F2 isoprostanes or malondialdehyde is commonly used to assess the extent of lipid peroxidation. Protein oxidation disrupts homeostasis by inactivating essential enzymes and altering the cell's ability to recognize and dispose of worn-out proteins. Scientists measure the level of protein carbonyls or advanced glycation end products to assess the amount of oxidative protein damage. Oxidative damage to nucleic acids can lead to deleterious mutations that may diminish cellular function or contribute to cancer development. The extent of oxidative lesions in mitochondrial DNA exceeds those seen in nuclear DNA by 10 to 1, suggesting that oxidative stress significantly contributes to disruption of mitochondrial function. The oxidative DNA adduct, 8-hydroxyguanosine, is the biochemical marker most widely used to measure the extent of oxidative DNA damage (Wu, Chiou, Chang, & Wu, 2004).

Cellular Antioxidant Defenses

Cellular protection from oxidative stress relies on enzymes that speed up ROS detoxification and on the activity of free radical scavengers. The antioxidant enzyme superoxide dismutase (SOD) catalyzes the conversion of superoxide anion to hydrogen peroxide; then hydrogen peroxide is converted to water by the enzymes catalase or glutathione peroxidase to complete the detoxification process. There are two distinct forms of SOD: (a) CuZnSOD, which is cytoplasmic and constitutively expressed and (b) MnSOD, which is mitochondrial and inducible. SOD activity is highly conserved across species, and interspecies comparisons show a strong positive correlation between SOD activity and maximum life span potential (Sohal, Sohal, & Brunk, 1990). Free radical scavengers that are hydrophilic, such as ascorbate, urate, and glutathione, operate within the cell cytoplasm. Lipophilic scavengers, such as carotenoids (e.g., beta carotene, lycopene) and tocopherols (e.g., vitamin E), provide valuable protection against peroxidation of membrane lipids.

Defining the precise role that a particular antioxidant defense enzyme plays in the aging process can be difficult for several reasons. First, some enzymes have sequential activity—SOD alone cannot drive the complete detoxification of superoxide anion to water. In addition, there is functional redundancy built into the system. Two different

enzymes, catalase and glutathione peroxidase, can complete the detoxification process of hydrogen peroxide to water. Furthermore, the inducibility of enzyme systems renders ambiguous the interpretation of “low” versus “high” measured enzyme activities. Should high basal SOD values be considered “good” or “bad”? The answer is a bit tricky. High activity might reflect innately strong defenses or, instead, induced activity suggesting the organism is under siege by overwhelming oxidative stress. Clearly, more studies are needed that not only measure baseline antioxidant enzyme activities but also assess cells *after oxidative challenge*.

Age-Related Changes in Oxidative Damage

In general, older hosts appear to be more susceptible to oxidative injury. For example, old mice exposed to low-dose (2Gy) irradiation had a twofold greater increase in oxidative DNA damage in the liver, brain, and heart than young mice receiving the same radiation exposure (Hamilton et al., 2001). In that study, evaluation of repair kinetics showed no difference in the capacity for young or old mice to repair oxidative DNA lesions (Hamilton et al., 2001). Studies probing whether age-related increases in oxidative damage reflect increased ROS production, diminished antioxidant defenses, or decreased repair of oxidative damage have yielded inconsistent results (Blumberg, 2000).

Interventional Studies

The evidence is mixed regarding whether manipulation of antioxidant defenses influences longevity. Transgenic flies that overexpress both CuZnSOD and catalase show increased maximum life span (Orr & Sohal, 1994). Similarly, selective overexpression of CuZnSOD in motor neurons also increases the maximum life span of flies (Parkes et al., 1998). However, upregulation of CuZnSOD activity in transgenic mice to levels up to five times greater than those seen in normal mice failed to increase life span (Huang, Carlson, Gillespie, Shi, & Epstein, 2000). Results from a more extensive catalog of studies in transgenic and knockout animals, in which a single gene or multiple genes have been overexpressed or deleted, have likewise failed to draw strong linkage between antioxidant defenses and life span in mammals (Salmon, Richardson, & Pérez, 2010).

Perhaps the most convincing evidence to support the oxidative stress hypothesis comes from the work of Melov and colleagues (2000) in *C. elegans*. Experimentally, the antioxidant defenses of worms were boosted by exposing them to a synthetic SOD mimetic in their culture

media, while control worms were exposed to vehicle alone. Worms exposed to the SOD mimetic had a significant 44% increase in mean life span. Furthermore, when genetically altered short-lived worms were exposed to the SOD mimetic, normal life span was restored in these short-lived mutants.

Unfortunately, direct evidence is lacking to support the hypothesis that increasing the dietary intake of antioxidants can increase life span in mammals. A synthetic free radical spin trap compound called PBN (N-tert-butyl alphaphenylnitrone) decreased brain protein oxidation (Carney et al., 1991) and increased life span of mice in some experiments (Floyd, Hensley, Forster, Kelleher-Andersson, & Wood, 2002). However, in studies of genetically heterogeneous mice, dietary supplementation with the PBN metabolite 4-hydroxy-PBN did not extend longevity (Strong et al., 2008). In humans, the dietary antioxidants vitamin C and vitamin E blocked the beneficial effects of exercise on blood glucose control, consistent with the notion that indiscriminant use of antioxidant supplements has the capacity to exert detrimental effects on human health (Ristow et al., 2009). Undoubtedly, new research will continue to challenge old thinking. And nowhere is this more clearly apparent than in the field of oxidative stress and aging.

LESSONS FROM THE OLDEST-OLD

Centenarians provide a potentially valuable natural experiment of highly successful aging. To date, the search for specific environmental exposures or lifestyle factors (e.g., diet, education, physical activity) that confer an increased likelihood of living to be 100 has not been fruitful (Perls, Levenson, Regan, & Puca, 2002). Among centenarians, females outnumber males by about four to one (Terry, Sebastiani, Anderson, & Perls, 2008). Among women, successful late reproduction—live childbirth at age 40 years or more—is a strong predictor of exceptional longevity (Perls, Alpert, & Frett, 1997). Larger particle size of high- and low-density lipoproteins (Barzilai et al., 2003) and higher plasma antioxidant capacity (Hyland et al., 2002) have also been associated with exceptional longevity.

Siblings of centenarians are 15 times more likely to live to be 100 compared with the general population (Perls, Wilmoth, et al., 2002). This longevity advantage is sustained throughout the life course, suggesting that their good fortune may be more strongly influenced by genetics than by environmental factors (Perls, Wilmoth, et al., 2002). A few genes have already been implicated as candidate longevity-enabling genes because the bad polymorphic variants of these genes are underrepresented in centenarians. These sinister variants

include the apolipoprotein Eε4 allele, and the angiotensin-converting enzyme D allele (Deelen et al., 2011; Schächter et al., 1994). Genetic linkage analysis has provided evidence that a gene or several genes on human chromosome 4 significantly contribute to exceptional longevity (Puca et al., 2001), while candidate gene studies point to two other longevity foci—FOXO3A and AKT1 (Christensen, Johnson, & Vaupel, 2006; Pawlikowska et al., 2009). It should be noted that the genetic factors that contribute to exceptional longevity may be different from the genetic determinants of average life span. In addition, men and women may follow different trajectories to achieve exceptional longevity; male centenarians are less heterogeneous and healthier than female centenarians (Franceschi, Motta, et al., 2000).

Centenarians outlive most of us, but just how healthy are centenarians? When it comes to vulnerability to major age-related diseases, a study of lifetime medical histories suggests there are three types of centenarians: (a) survivors (38%)—individuals with onset of at least one major disease prior to 80 years of age; (b) delayers (43%)—individuals who are free of all major diseases until after 80 years of age; and (c) escapers (19%)—individuals who are free of all major diseases until after 100 years of age (Evert, Lawler, Bogan, & Perls, 2003). This important study confirmed that centenarians are indeed a highly heterogeneous population. It is apparent that strong disease resistance, however, is a characteristic shared by the majority of centenarians; almost two-thirds of centenarians remained free of all major diseases until after the median age at death for their birth cohort. An analysis of exceptional longevity in pet dogs reached a similar conclusion: 76% of oldest-old dogs delayed the onset of major diseases, and more than one-half of extreme-aged dogs had profound disease resistance (Cooley, Schlittler, Glickman, Hayek, & Waters, 2003). In humans, mortality from cancer, a disease strongly associated with aging, actually *declines* in the tenth decade of life—a puzzling paradox (Harding, Pompei, & Wilson, 2011; Smith, 1996; Stanta, Campagner, Cavallieri, & Giarelli, 1997). Like the oldest-old humans, extreme-aged pet dogs also experience a significant decline in cancer-related mortality (Cooley et al., 2003; Perls & Hutter-Silver, 1999).

Several aspects of studying human centenarians present problematic hurdles for researchers: age validation, stochastic influences, concurrent disease, selection of adequate controls for comparison, and long-term recall of diet and other lifestyle factors. Recognizing the big payoff that could come from deciphering some of the secrets behind highly successful aging, biogerontologists have been on the lookout for fresh, new approaches that would be complementary to studies of human centenarians. In 2005, we established the Center for Exceptional Longevity Studies at the Gerald P. Murphy Cancer

Foundation. The aim was to test a new idea: The secrets to successful human aging might be revealed by studying exceptional longevity in pet dogs. To accomplish this, we set out to conduct the first, systematic study of the oldest-old dogs, starting with Rottweilers. We created ELDABA (Exceptional Longevity DataBase), the world's first collection of medical history and lifestyle information on canine centenarians (for Rottweilers, this means still kicking at the age of 13). This work has already led to important discoveries. By carefully studying the association between the number of years of lifetime ovary exposure and longevity in female Rottweilers, we discovered that keeping ovaries longer is associated with living longer (Waters et al., 2009). This work in dogs, together with findings on the longevity-promoting effects of ovaries in women and mice point to a new line of thinking: Ovaries are part of a system that promotes longevity (Mason, Cargill, Anderson, & Carey, 2009; Parker et al., 2009; Rocca, Grossardt, de Andrade, Malkasian, & Melton, 2006; Waters et al., 2009).

APPLYING A LIFE COURSE PERSPECTIVE TO BIOGERONTOLOGICAL DISCOVERY

Biogerontologists should not confine themselves to the study of old cells, old dogs, or old people. Instead, it makes better sense to apply a life course perspective to studying the determinants of healthy longevity, because the origins of many adult health outcomes are shaped significantly by early-life events. For example, the risk of older people to suffer a pathologic bone fracture secondary to osteoporosis strongly depends on the peak bone mass established in the second and third decades of life.

The discovery of biomarkers that can accurately identify those individuals in a population that are aging at different rates is a high research priority. Finding reliable biomarkers of aging, however, has been a frustrating failure, forcing investigators to fall back on longevity as the end point for their studies testing antiaging interventions. Could paying closer attention to life course perspective go far to fit together some of the pieces of the biomarker puzzle? We think so. It turns out that the relationship between certain physiologic parameters and health outcomes in people apparently “flip-flops” during the life course. For example, high serum cholesterol at 50 years of age is associated with an increased risk of developing Alzheimer's disease 20 years later (Kivipelto et al., 2001). In contrast, in people who are 70 years of age, high cholesterol seems to be *protective* against Alzheimer's disease (van Vliet, van de Water, de Craen, & Westendorp, 2009; Zuliani et al., 2010). Human studies likewise suggest that obesity in young

and middle-aged populations is associated with diminished life expectancy, whereas in old age (beyond 60 years, especially in men), fatness loses its sting as a harbinger of earlier death (Freedman, Ron, Ballard-Barbash, Doody, & Linet, 2006; Thinggaard, Jacobsen, Jeune, Martinussen, & Christensen, 2010). In advanced age, more favorable survival may be more closely related to increasing adiposity than muscle mass (Auyeung et al., 2010).

So how might we reconcile these puzzling observations that seem to be leading us toward a less certain view of how we can best promote healthy longevity? Instead of seeing the aging organism as engaged in a monotonous, downward trajectory, we need to see the organism as a dynamic and changing system, capable of resetting itself during the aging process to remain as resilient as possible amid the challenges and constraints of physiologic decline. With this reality more clearly in view, we should reexamine carefully not only our choices of interventions designed to sidestep the disabilities of aging, but also examine how we might *optimize the timing of those interventions*. It is expected that age at the start of any intervention, such as a CR mimetic, will strongly influence that intervention's longevity-altering effect. In some instances, earlier intervention will be preferred over late. It is likely that some interventions may be specific for promoting or restoring health only later in life. Importantly, we will need to look closely for the tradeoffs that will inevitably occur between beneficial and detrimental effects, so we can intelligently optimize our choices.

THE FRONTIER OF BIOGERONTOLOGICAL DISCOVERY: CRITICAL KNOWLEDGE GAPS, FAR-REACHING OPPORTUNITIES

The next generation of biogerontologists must be transdisciplinary who excel in their home discipline yet are conversant in related areas of inquiry. Amid the substantial advances in our understanding of the aging process, we see four critical gaps that translate into opportunities:

1. *There is a critical need to identify biomarkers that can accurately identify individuals who are aging at significantly different rates.* Much recent attention has focused on biomarkers of inflammation. A decade ago, Franceschi, Bonafè, and colleagues (2000) coined the term *inflammaging* to describe the apparent pro-inflammatory state of elderly humans. Since then, numerous studies have documented elevated levels of inflammatory cytokines, such as IL-6 and TNF-alpha, in older adults fueling the

hypothesis that aging is innately pro-inflammatory (Danesh et al., 2008). However, other investigators have challenged whether increased inflammation is an obligate phenotype, positing that the pro-inflammatory state of the elderly may reflect an accumulation of lifestyle factors, particularly obesity, or the high incidence of atherosclerosis and other comorbid diseases (Ferrucci et al., 2005). Validation of biomarkers is an essential step toward the rigorous evaluation of interventions to slow the rate of senescence (Nakamura, Lane, Roth, & Ingram, 1998). Without them, investigators will have to rely on life span as an end point—making studies in long-lived species like humans impractical. Moreover, in the absence of validated biomarkers of senescence, the public will continue to be bombarded by unsubstantiated claims from peddlers of antiaging interventions.

2. *There is genuine concern about the relevance of the model systems used to study aging.* At best, animal and cellular models are imitations of human aging. It is imperative that investigators understand and freely communicate the strengths and limitations of the experimental model systems they use. Animal models should never drive the research questions. Instead, the particular research question should determine the selection of the most appropriate animal model (Waters, Bostwick, & Murphy, 1998). There is a recognized need for development of a broader array of model systems, including birds (Holmes & Austad, 1995), marsupials (Austad, 1997), and other mammals (Miller et al., 1999), to test important hypotheses relevant to human aging. A recent issue of *The ILAR Journal* was devoted to benchmarking current thinking about animal models of human health span (Sprott, 2011; Waters, 2011).
3. *There is need for a coherent theoretical framework that can be used to test important hypotheses.* Useful conceptual models should distinguish between senescence and longevity. Aging is complex and the process of senescence is undoubtedly driven by multiple causal mechanisms. It seems rational therefore to apply systems thinking so that we might integrate key concepts and side-step the intellectual paralysis potentially produced by the vast number of competing theories, now numbered in excess of 300 (Kirkwood, 2005; Medvedev, 1990). Development of a consensus framework would undoubtedly help investigators posit hypotheses and propose experiments to answer the most fundamental of all questions: *Why do old tissues have increased vulnerability to disease?* This should also enable the formal testing of the hypothesis that a slower rate of aging is, in fact, an important determinant of exceptional longevity.

4. *It makes good sense to apply a life course perspective to studying the determinants of life span.* Biogerontology should not be confined to the study of old organisms because the origins of many adult health outcomes are shaped significantly by early life events and experiences. A contemporary approach to finding the determinants of successful aging must also integrate how psychosocial factors modify physiologic homeostasis and the trajectory of age-related functional decline. We must begin to analyze the extent to which the timing and sequence, as well as the duration and intensity of experiences and exposures impact aging trajectories. The wisdom of using this kind of integrative approach is illustrated by the groundbreaking work documenting that the number of adverse childhood events has a significant impact on adult health outcomes (Brown et al., 2010; Brown et al., 2009; Felitti et al., 1998). It follows that careful studies of just how epigenetic reprogramming during the life course can be exploited to prevent or reverse environmentally driven, adverse health outcomes should command high priority (Weaver et al., 2006).

REFERENCES

- Aguilera, O., Fernández, A. F., Muñoz, A., & Fraga, M. F. (2010). Epigenetics and environment: A complex relationship. *Journal of Applied Physiology*, *109*, 243–251.
- Anson, R. M., Guo, Z., de Cabo, R., Iyun, T., Rios, M., Hagepanos, A., ... Mattson, M. (2003). Intermittent fasting dissociates beneficial effects of dietary restriction on glucose metabolism and neuronal resistance to injury from calorie intake. *Proceedings of the National Academy of Sciences of the United States of America*, *100*, 6216–6220.
- Arking, R. (1987). Successful selection for increased longevity in *Drosophila*: Analysis of the survival data and presentation of a hypothesis on the genetic regulation of longevity. *Experimental Gerontology*, *22*, 199–220.
- Austad, S. N. (1993). Retarded senescence in an insular population of Virginia opossums. *Journal of Zoology (London)*, *229*, 695–708.
- Austad, S. N. (1997). Comparative aging and life histories in mammals. *Experimental Gerontology*, *32*, 23–38.
- Auyeung, T. W., Lee, J. S., Leung, J., Kwok, T., Leung, P. C., & Woo, J. (2010). Survival in older men may benefit from being slightly overweight and centrally obese—a 5-year follow-up study in 4,000 older adults using DXA. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *65*, 99–104.
- Bakaysa, S. L., Mucci, L. A., Slagboom, P. E., Boomsma, D. I., McClearn, G. E., Johansson, B., & Pedersen, N. L. (2007). Telomere length predicts survival independent of genetic influences. *Aging Cell*, *6*, 769–774.
- Bartke, A. (2008). Impact of reduced insulin-like growth factor-1/insulin signaling on aging in mammals: Novel findings. *Aging Cell*, *7*, 285–290.

- Bartke, A., Brown-Borg, H. M., Bode, A. M., Carlson, J., Hunter, W. S., & Bronson, R. T. (1998). Does growth hormone prevent or accelerate aging? *Experimental Gerontology*, *33*, 675–687.
- Barzilai, N., Atzmon, G., Schechter, C., Schaefer, E. J., Cupples, A. L., Lipton, R., ... Shuldiner, A. (2003). Unique lipoprotein phenotype and genotype associated with exceptional longevity. *Journal of the American Medical Association*, *290*, 2030–2040.
- Beckman, K. B., & Ames, B. N. (1998). The free radical theory of aging matures. *Physiological Reviews*, *78*, 547–581.
- Bhasin, S., Cunningham, G. R., Hayes, F. J., Matsumoto, A. M., Snyder, P. J., Swerdloff, R. S., & Montori, V. M. (2010). Testosterone therapy in men with androgen deficiency syndromes: An endocrine society clinical practice guideline. *Journal of Clinical Endocrinology and Metabolism*, *95*, 2536–2559.
- Binstock, R. H. (2004). Anti-aging medicine and research: A realm of conflict and profound societal implications. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *59*, B523–B533.
- Blackman, M. R., Sorkin, J. D., Münzer, T., Bellantoni, M. F., Busby-Whitehead, J., Stevens, T. E., ... Harman, S. M. (2002). Growth hormone and sex steroid administration in healthy aged women and men: A randomized controlled trial. *Journal of the American Medical Association*, *288*, 2282–2292.
- Blumberg, J. B. (2000). Free radical theory of aging. In J. E. Morley, H. J. Armbrecht, R. M. Coe, & B. Vellas (Eds.), *The science of geriatrics* (Vol. 1, pp. 57–74). New York, NY: Springer Publishing.
- Brown, D. W., Anda, R. F., Felitti, V. J., Edwards, V. J., Malarcher, A. M., Croft, J. B., & Giles, W. H. (2010). Adverse childhood experiences are associated with the risk of lung cancer: A prospective cohort study. *BMC Public Health*, *10*, 20.
- Brown, D. W., Anda, R. F., Tiemeier, H., Felitti, V. J., Edwards, V. J., Croft, J. B., Giles, W. H. (2009). Adverse childhood experiences and the risk of premature mortality. *American Journal of Preventive Medicine*, *37*, 389–396.
- Butler, R. N., Austad, S. N., Barzilai, N., Braun, A., Helfand, S., Larsen, P. L., ... Warner, H. R. (2003). Longevity genes: From primitive organisms to humans. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *58*, 581–584.
- Campisi, J. (1996). Replicative senescence: An old lives' tale? *Cell*, *84*, 497–500.
- Carney, J. M., Starke-Reed, P. E., Oliver, C. N., Landum, R. W., Cheng, M. S., Wu, J. F., & Floyd, R. A. (1991). Reversal of age-related increase in brain protein oxidation, decrease in enzyme activity, and loss in temporal and spatial memory by chronic administration of the spin-trapping compound N-tertbutyl-alpha-phenylnitron. *Proceedings of the National Academy of Sciences of the United States of America*, *88*, 3633–3636.
- Chan, J. M., Stampfer, M. J., Giovannucci, E., Gann, P. H., Ma, J., Wilkinson, P., ... Pollak, M. (1998). Plasma insulin-like growth factor-I and prostate cancer risk: A prospective study. *Science*, *279*, 563–566.
- Chiang, E. C., Shen, S., Kengeri, S. S., Xu, H., Combs, G. F., Jr., Morris, J. S., ... Waters, D. J. (2010). Defining the optimal dose of selenium for prostate

- cancer risk reduction: Insights from the U-shaped relationship between selenium status, DNA damage, and apoptosis. *Dose Response*, 8, 285–300.
- Christensen, K., Johnson, T. E., & Vaupel, J. W. (2006). The quest for genetic determinants of human longevity. *Nature Reviews. Genetics*, 7, 436–448.
- Clancy, D. J., Gems, D., Harshman, L. G., Oldham, S., Stocker, H., Hafen, E., ... Partridge, L. (2001). Extension of life span by loss of CHICO, a Drosophila insulin receptor substrate protein. *Science*, 292, 104–106.
- Colman, R. J., Anderson, R. M., Johnson, S. C., Kastman, E. K., Kosmatka, K. J., Beasley, T. M. ... Weindruch, R. (2009). Caloric restriction delays disease onset and mortality in rhesus monkeys. *Science*, 325, 201–204.
- Cooley, D. M., Schlittler, D. L., Glickman, L. T., Hayek, M., & Waters, D. J. (2003). Exceptional longevity in pet dogs is accompanied by cancer resistance and delayed onset of major diseases. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, 58, B1078–B1084.
- Cristofalo, V. J., Allen, R. G., Pignolo, R. J., Martin, B. G., & Beck, J. C. (1998). Relationship between donor age and the replicative life span of human cells in culture: A reevaluation. *Proceedings of the National Academy of Sciences of the United States of America*, 95, 10614–10619.
- Cristofalo, V. J., & Pignolo, R. J. (1993). Replicative senescence of human fibroblast-like cells in culture. *Physiological Reviews*, 73, 617–638.
- Cuervo, A. M. (2010). Chaperone-mediated autophagy: Selectivity pays off. *Trends in Endocrinology and Metabolism*, 21, 142–150.
- Danesh, J., Kaptoge, S., Mann, A. G., Sarwar, N., Wood, A., Angleman, S. B., ... Gudnason, V. (2008). Long-term interleukin-6 levels and subsequent risk of coronary heart disease: Two new prospective studies and a systematic review. *PLoS Medicine*, 5, e78.
- Davalos, A., Coppe, J., Campisi, J., & Desprez P. (2010). Senescent cells as a source of inflammatory factors for tumor progression. *Cancer Metastasis Reviews*, 29, 273–283.
- Debacq-Chainiaux, F., Erusalimsky, J. D., Campisi, J., & Toussaint, O. (2009). Protocols to detect senescence-associated beta-galactosidase (SA-beta-gal) activity, a biomarker of senescent cells in culture and in vivo. *Nature Protocols*, 4, 1798–1806.
- Deeb, B. J., & Wolf, N. S. (1994). Studying longevity and morbidity in giant and small breeds of dogs. *Veterinary Medicine (Suppl.)*, 89, 702–713.
- Deelen, J., Beekman, M., Uh, H., Helmer, Q., Kuningas, M., Christiansen, L., ... Slagboom, P. (2011). Genome-wide association study identifies a single major locus contributing to survival into old age; the *APOE* locus revisited. *Aging Cell*, 10, 686–698.
- Dekker, P., Maier, A. B., van Heemst, D., de Koning-Treurniet, C., Blom, J., Dirks, R. W., ... Westendorp, R. G. (2009). Stress-induced responses of human skin fibroblasts *in vitro* reflect human longevity. *Aging Cell*, 8, 595–603.
- Dimri, G. (2005). What has senescence got to do with cancer? *Cancer Cell*, 7, 505–512.
- Dimri, G. P., Lee, X., Basile, G., Acosta, M., Scott, G., Roskelley, C., ... Pereira-Smith, O. (1995). A biomarker that identifies senescent human cells in culture and in aging skin in vivo. *Proceedings of the National Academy of Sciences of the United States of America*, 92, 9363–9367.

- Epel, E. S., Blackburn, E. H., Lin, J., Dhabhar, F. S., Adler, N. E., Morrow, J. D., & Cawthon, R. M. (2004). Accelerated telomere shortening in response to life stress. *Proceedings of the National Academy of Sciences of the United States of America*, *101*, 17312–17315.
- Evert, J., Lawler, E., Bogan, H., & Perls, T. (2003). Morbidity profiles of centenarians: Survivors, delayers, and escapers. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *58*, 232–237.
- Felitti, V. J., Anda, R. F., Nordenberg, D., Williamson, D. F., Spitz, A. M., Edwards, V., ... Marks, J. S. (1998). Relationship of childhood abuse and household dysfunction to many of the leading causes of death in adults. The Adverse Childhood Experiences (ACE) Study. *American Journal of Preventive Medicine*, *14*, 245–258.
- Ferrucci, L., Corsi, A., Lauretani, F., Bandinelli, S., Bartali, B., Taub, D. D., ... Longo, D. (2005). The origins of age-related proinflammatory state. *Blood*, *105*, 2294–2299.
- Finch, C. E. (1990). *Longevity, senescence and the genome*. Chicago, IL: University of Chicago Press.
- Finch, C. E. (1998). Variations in senescence and longevity include the possibility of negligible senescence. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *53A*, 232–237.
- Finch, C. E. (2009). Update of slow aging and negligible senescence—a mini-review. *Gerontology*, *55*, 307–313.
- Finch, C. E., & Austad, S. N. (2001). History and prospects: Symposium on organisms with slow aging. *Experimental Gerontology*, *36*, 593–597.
- Finch, C. E., & Kirkwood, T. B. L. (2000). *Chance, development and aging*. New York, NY: Oxford University Press.
- Finch, C. E., Morgan, T. E., Longo, V. D., & de Magalhaes, J. P. (2010). Cell resilience in species life spans: A link to inflammation? *Aging Cell*, *9*, 519–526.
- Floyd, R. A., Hensley, K., Forster, M. J., Kelleher-Andersson, J. A., & Wood, P. L. (2002). Nitrones, their value as therapeutics and probes to understand aging. *Mechanisms of Ageing and Development*, *123*, 1021–1031.
- Franceschi, C., Bonafè, M., Valensin, S., Olivieri, F., De Luca, M., Ottaviani E., & De Benedictis, G. (2000). Inflamm-aging. An evolutionary perspective on immunosenescence. *Annals of the New York Academy of Sciences*, *908*, 244–254.
- Franceschi, C., Motta, L., Valensin, S., Rapisarda, R., Franzone, A., Berardelli, M., ... Baggio, G. (2000). Do men and women follow different trajectories to reach extreme longevity? *Aging (Milano)*, *12*, 77–84.
- Freedman, D. M., Ron, E., Ballard-Barbash, R., Doody, M. M., & Linet, M. S. (2006). Body mass index and all-cause mortality in a nationwide US cohort. *International Journal of Obesity*, *30*, 822–829.
- Fries, J. F. (2003). Measuring and monitoring success in compressing morbidity. *Annals of Internal Medicine*, *139*, 455–459.
- Giovannucci, E., Pollak, M. N., Platz, E. A., Willett, W. C., Stampfer, M. J., Majeed, N., ... Hankinson, S. E. (2000). A prospective study of plasma insulin-like growth factor-1 and binding protein-3 and risk of colorectal neoplasia in women. *Cancer Epidemiology, Biomarkers and Prevention*, *9*, 345–349.

- Grandison, R.C., Piper, M. D., & Partridge, L. (2009). Amino acid imbalance explains extension of life span by dietary restriction in *Drosophila*. *Nature*, *462*, 1061–1064.
- Granger, M. P., Wright, W. E., & Shay, J. W. (2002). Telomerase in cancer and aging. *Critical Reviews in Oncology/Hematology*, *41*, 29–40.
- Gravina, S., & Vijg, J. (2010). Epigenetic factors in aging and longevity. *Pflügers Arch: European Journal of Physiology*, *459*, 247–258.
- Hamilton, M. L., Van Remmen, H., Drake, J. A., Yang, H., Guo, Z. M., Kewitt, K., ... Richardson, A. (2001). Does oxidative damage to DNA increase with age? *Proceedings of the National Academy of Sciences of the United States of America*, *98*, 10469–10474.
- Harding, C., Pompei, F., & Wilson, R. (2011). Peak and decline in cancer incidence, mortality, and prevalence at old ages. *Cancer*, *118*, 1371–1386. DOI: 10.1002/cncr.26376
- Harman, D. (1956). Aging: A theory based on free radical and radiation chemistry. *Journal of Gerontology*, *11*, 298–300.
- Harper, J. M., Leathers, C. W., & Austad, S. N. (2006). Does caloric restriction extend life in wild mice? *Aging Cell*, *5*, 441–449.
- Harper, J. M., Salmon, A. B., Leiser, S. F., Galecki, A. T., & Miller, R. A. (2007). Skin-derived fibroblasts from long-lived species are resistant to some, but not all, lethal stresses and to the mitochondrial inhibitor rotenone. *Aging Cell*, *6*, 1–13.
- Hass, B. S., Lewis, S. M., Duffy, P. H., Ershler, W., Feuers, R. J., Good, R. A., ... Hart, R. W. (1996). Dietary restriction in humans: Report on the Little Rock Conference on the value, feasibility, and parameters of a proposed study. *Mechanisms of Ageing and Development*, *91*, 79–94.
- Hayflick, L. (1976). The cell biology of human aging. *New England Journal of Medicine*, *295*, 1302–1308.
- Hayflick, L. (1998). How and why we age. *Experimental Gerontology*, *33*, 639–653.
- Hayflick, L., & Moorhead, P. S. (1961). The serial cultivation of human diploid cell strains. *Experimental Cell Research*, *25*, 585–621.
- Hodes, R., McCormick, A., & Pruzan, M. (1996). Longevity assurance genes: How do they influence aging and life span? *Journal American Geriatric Society*, *44*, 988–991.
- Holmes, D. J., & Austad, S. N. (1995). Birds as animal models for the comparative biology of aging: A prospectus. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *50*, B59–B66.
- Huang, T., Carlson, E., Gillespie, A., Shi, Y., & Epstein, C. (2000). Ubiquitous over-expression of CuZn superoxide dismutase does not extend life span in mice. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *55*, B5–B9.
- Hyland, P., Duggan, O., Turbitt, J., Coulter, J., Wikby, A., Johansson, B., ... Barnett, Y. (2002). Nonagenarians from the Swedish NONA Immune Study have increased plasma antioxidant capacity and similar levels of DNA damage in peripheral blood mononuclear cells compared to younger control subjects. *Experimental Gerontology*, *37*, 465–473.

- Jordan, V. C. (2004). Selective estrogen receptor modulation: Concept and consequences in cancer. *Cancer Cell*, *5*, 207–213.
- Kim Sh, S. H., Kaminker, P., & Campisi, J. (2002). Telomeres, aging and cancer: In search of a happy ending. *Oncogene*, *21*, 503–511.
- Kimura, K. D., Tissenbaum, H. A., Liu, Y., & Ruvkun, G. (1997). *daf-2*, an insulin receptor-like gene that regulates longevity and diapause in *Caenorhabditis elegans*. *Science*, *277*, 942–946.
- Kirkwood, T. B. (1977). Evolution of ageing. *Nature*, *270*, 301–304.
- Kirkwood, T. B. (1990). The disposable soma theory of aging. In D. E. Harrison, (Ed.), *Genetic effects on aging* (Vol. 2, pp. 9–19). Caldwell, NJ: Telford Press.
- Kirkwood, T. B. L. (2005). Understanding the odd science of aging. *Cell*, *120*, 437–447.
- Kivipelto, M., Helkala, E-L., Laakso, M. P., Hänninen, T., Hallikainen, M., Alhainen, K., ... Nissinen, A. (2001). Midlife vascular risk factors and Alzheimer's disease in later life: Longitudinal, population based study. *British Medical Journal*, *322*, 1447–1451.
- Kizer, J., Arnold, A., Jenny, N., Cushman, M., Strotmeyer, E., Ives, D., ... Newman, A. (2011). Longitudinal changes in adiponectin and inflammatory markers and relation to survival in the oldest old: The cardiovascular health study all stars study. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *66*, 1100–1107.
- Koga, H., Kaushik, S., & Cuervo, A. M. (2011). Protein homeostasis and aging: The importance of exquisite quality control. *Aging Research Reviews*, *10*, 205–215.
- Krause, W., Mueller, U., & Mazur, A. (2005). Testosterone supplementation in the aging male: Which questions have been answered? *Aging Male*, *8*, 31–38.
- Lamberts, S. W., van den Beld, A. W., & van der Lely, A. J. (1997). The endocrinology of aging. *Science*, *278*, 419–424.
- Lee, B., Han, J., Im, J., Morrone, A., Johung, K., Goodwin, E., ... Hwang, E. (2006). Senescence-associated β -galactosidase is lysosomal β -galactosidase. *Aging Cell*, *5*, 187–195.
- Liao, C. Y., Rikke, B. A., Johnson, T. E., Diaz, V., & Nelson, J. F. (2010). Genetic variation in the murine life span response to dietary restriction: From life extension to life shortening. *Aging Cell*, *9*, 92–95.
- Liao, C. Y., Rikke, B. A., Johnson, T. E., Gelfond, J. A. L., Diaz, V., & Nelson, J. F. (2011). Fat maintenance is a predictor of the murine life span response to dietary restriction. *Aging Cell*, *10*, 629–639.
- Lin, Y. J., Seroude, L., & Benzer, S. (1998). Extended life span and stress resistance in the *Drosophila* mutant methuselah. *Science*, *282*, 943–946.
- Ljungquist, B., Berg, S., Lanke, J., McClearn, G. E., & Pedersen, N. L. (1998). The effect of genetic factors for longevity: A comparison of identical and fraternal twins in the Swedish Twin Registry. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *53*, M441–M446.
- Love, R. R., Mazess, R. B., Barden, H. S., Epstein, S., Newcomb, P. A., Jordan, V. C., ... DeMets, D. L. (1992). Effects of tamoxifen on bone mineral density in postmenopausal women with breast cancer. *New England Journal of Medicine*, *326*, 852–856.

- Maier, A. B., Cessie, S., Koning-Treurniet, C., Blom, J., Westendorp, R. G. J., & Heemst, D. (2007). Persistence of high-replicative capacity in cultured fibroblasts from nonagenarians. *Aging Cell*, *6*, 27–33.
- Martin, B., Pearson, M., Brenneman, R., Golden, E., Keselman, A., Iyun, T., ... Mattson, M. P. (2008). Conserved and differential effects of dietary energy intake on the hippocampal transcriptomes of females and males. *PLoS One*, *3*, e2398.
- Martin, B., Pearson, M., Brenneman, R., Golden, E., Wood III, W., Prabhu, V., ... Maudsley, S. (2009). Gonadal transcriptome alterations in response to dietary energy intake: Sensing the reproductive environment. *PLoS One*, *4*, e4146.
- Martin, G. M. (1978). Genetic syndromes in man with potential relevance to the pathobiology of aging. In D. Bergsma & D. E. Harrison (Eds.), *Genetic effects on aging* (Vol. 14, pp. 5–39). New York, NY: Alan R. Liss.
- Martin, G. M., Austad, S. N., & Johnson, T. E. (1996). Genetic analysis of ageing: Role of oxidative damage and environmental stresses. *Nature Genetics*, *13*, 25–34.
- Martin, G., Sprague, C., & Epstein, C. (1970). Replicative life span of cultivated human cells. Effects of donor's age, tissue, and genotype. *Lab Investigation*, *23*, 86–92.
- Mason J. B., Cargill, S. L., Anderson, G. B., & Carey, J. R. (2009). Transplantation of young ovaries to old mice increased life span in transplant recipients. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *64*, 1207–1211.
- Masoro E. J. (2003). Subfield history: Caloric restriction, slowing aging, and extending life. *Scientific Aging Knowledge Environment*, *8*, RE2. Review.
- Mather, K. A., Jorm, A. F., Parslow, R. A., & Christensen, H. (2011). Is telomere length a biomarker of aging? A review. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *66*, 203–213.
- McCay, C., Crowell, M., & Maynard, L. (1935). The effect of retarded growth upon the length of life span and upon the ultimate body size. *Journal of Nutrition*, *10*, 63–79.
- McGue, M., Vaupel, J. W., Holm, N., & Harvald, B. (1993). Longevity is moderately heritable in a sample of Danish twins born 1870–1880. *Journal of Gerontology*, *48*, B237–B244.
- Medvedev, Z. A., (1990). An attempt at a rational classification of theories of aging. *Biological Reviews of the Cambridge Philosophical Society*, *65*, 375–398.
- Melov, S., Ravenscroft, J., Malik, S., Gill, M. S., Walker, D. W., Clayton, P. E., ... Lithgow, G. J. (2000). Extension of life span with superoxide dismutase/catalase mimetics. *Science*, *289*, 1567–1569.
- Migliaccio, E., Giorgio, M., Mele, S., Pelicci, G., Reboldi, P., Pandolfi, P. P., ... Pelicci, P. G. (1999). The p66shc adaptor protein controls oxidative stress response and life span in mammals. *Nature*, *402*, 309–313.
- Miller, R. A., Austad, S., Burke, D., Chrisp, C., Dysko, R., Galecki, A., ... Monnier, V. (1999). Exotic mice as models for aging research: Polemic and prospectus. *Neurobiology of Aging*, *20*, 217–231.

- Miller, R. A., Williams, J. B., Kiklevich, J. V., Austad, S., & Harper, J. M. (2011). Comparative cellular biogerontology: Primer and prospectus. *Ageing Research Reviews, 10*, 181–190.
- Miquel, J. (1992). An update on the mitochondrial-DNA mutation hypothesis of cell aging. *Mutation Research, 275*, 209–216.
- Monteiro, H., & Stern, A. (1996). Redox modulation of tyrosine phosphorylation-dependent signal transduction pathways. *Free Radical Biology Medicine, 21*, 323–333.
- Morris, J. Z., Tissenbaum, H. A., & Ruvkun, G. (1996). A phosphatidylinositol-3-OH kinase family member regulating longevity and diapause in *Caenorhabditis elegans*. *Nature, 382*, 536–539.
- Munhoz, C., Sorrells, S., Caso, J., Scavone, C., & Sapolsky, R. (2010). Glucocorticoids exacerbate lipopolysaccharide-induced signaling in the frontal cortex and hippocampus in a dose-dependent manner. *Journal of Neuroscience, 30*, 13690–13698.
- Nakamura, E., Lane, M. A., Roth, G. S., & Ingram, D. K. (1998). A strategy for identifying biomarkers of aging: Further evaluation of hematology and blood chemistry data from a calorie restriction study in rhesus monkeys. *Experimental Gerontology, 33*, 421–443.
- Olshansky, S. J., Hayflick, L., & Carnes, B. A. (2002). Position statement on human aging. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences, 57*, B292–B297.
- Orr, W., & Sohal, R. (1994). Extension of life span by overexpression of superoxide dismutase and catalase in *Drosophila melanogaster*. *Science, 263*, 1128–1130.
- Parker, W. H., Broder, M. S., Chang, E., Feskanich, D., Farguhar, C., Liu, Z., ... Manson, J. (2009). Ovarian conservation at the time of hysterectomy and long-term health outcomes in the Nurses' Health Study. *Obstetrics and Gynecology, 113*, 1027–1037.
- Parkes, T. L., Elia, A. J., Dickinson, D., Hilliker, A. J., Phillips, J. P., & Boulianne, G. L. (1998). Extension of *Drosophila* life span by overexpression of human SOD1 in motoneurons. *Nature Genetics, 19*, 171–174.
- Patronek, G. J., Waters, D. J., & Glickman, L. T. (1997). Comparative longevity of pet dogs and humans: Implications for gerontology research. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences, 52*, B171–B178.
- Pawlikowska, L., Hu, D., Huntsman, S., Sung, A., Chu, C., Chen, J., ... Study of Osteoporotic Fractures. (2009). Association of common genetic variation in the insulin/IGF1 signaling pathway with human longevity. *Ageing Cell, 8*, 460–472.
- Pérez, V. I., Buffenstein, R., Masamsetti, V., Leonard, S., Salmon, A. B., Mele, J., ... Chaudhuri, A. (2009). Protein stability and resistance to oxidative stress are determinants of longevity in the longest-living rodent, the naked mole-rat. *Proceedings of the National Academy of Sciences of the United States of America, 106*, 3059–3064.
- Perls, T., Alpert, L., & Frett, R. (1997). Middle aged mothers live longer. *Nature, 389*, 133.

- Perls, T., & Hutter-Silver, M. (1999). *Living to 100: Lessons in living to your maximum potential at any age*. New York, NY: Basic Books.
- Perls, T., Levenson, R., Regan, M., & Puca, A. (2002). What does it take to live to 100? *Mechanisms of Ageing and Development*, *123*, 231–242.
- Perls, T. T., Wilmoth, J., Levenson, R., Drinkwater, M., Cohen, M., Bogan, H., ... Puca, A. (2002). Life-long sustained mortality advantage of siblings of centenarians. *Proceedings of the National Academy of Sciences of the United States of America*, *99*, 8442–8447.
- Piper, M. D., Partridge, L., Raubenheimer, D., & Simpson, S. J. (2011). Dietary restriction and aging: A unifying perspective. *Cell Metabolism*, *14*, 154–160.
- Piper, M. D., Selman, C., McElwee, J. J., & Partridge, L. (2008). Separating cause from effect: How does insulin/IGF signalling control life span in worms, flies, and mice? *Journal of Internal Medicine*, *263*, 179–191.
- Powers, E. T., Morimoto, R. I., Dillin, A., Kelly, J. W., & Balch, W. E. (2009). Biological and chemical approaches to diseases of proteostasis deficiency. *Annual Review of Biochemistry*, *78*, 959–991.
- Puca, A. A., Daly, M. J., Brewster, S. J., Matise, T. C., Barrett, J., Shea-Drinkwater, M., ... Perls, T. T. (2001). A genome-wide scan for linkage to human exceptional longevity identifies a locus on chromosome 4. *Proceedings of the National Academy of Sciences of the United States of America*, *98*, 10505–10508.
- Pugh, T. D., Oberley, T. D., & Weindruch, R. (1999). Dietary intervention at middle age: Caloric restriction but not dehydroepiandrosterone sulfate increases life span and lifetime cancer incidence in mice. *Cancer Research*, *59*, 1642–1648.
- Ramsey, M., Ingram, R., Cashion, A., Ng, A., Cline, J., Parlow, A., & Sonntag, W. (2002). Growth hormone-deficient dwarf animals are resistant to dimethylbenzanthracene (DMBA)-induced mammary carcinogenesis. *Endocrinology*, *143*, 4139–4142.
- Rea, S. L., Wu, D., Cypser, J. R., Vaupel, J. W., & Johnson, T. E. (2005). A stress-sensitive reporter predicts longevity in isogenic populations of *Caenorhabditis elegans*. *Nature Genetics*, *37*, 894–898.
- Ristow, M., Zarse, K., Oberbach, A., Klötting, N., Birringer, M., Kiehnopf, M., ... Blüher, M. (2009). Antioxidants prevent health-promoting effects of physical exercise in humans. *Proceedings of the National Academy of Sciences of the United States of America*, *106*, 8665–8670.
- Roberts, S. B., Pi-Sunyer, X., Kuller, L., Lane, M. A., Ellison, P., Prior, J. C., & Shapses, S. (2001). Physiologic effects of lowering caloric intake in non-human primates and nonobese humans. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *56*, 66–75.
- Rocca, W. A., Grossardt, B. R., de Andrade, M., Malkasian, G. D., & Melton III, L. J. (2006). Survival patterns after oophorectomy in postmenopausal women: A population-based cohort study. *Lancet Oncology*, *7*, 821–828.
- Rodier, F., & Campisi, J. (2011). Four faces of cellular senescence. *Journal of Cell Biology*, *192*, 547–556.
- Rudman, D., Feller, A. G., Nagraj, H. S., Gergans, G. A., Lalitha, P. Y., Goldberg, A. F., ... Mattson, D. E. (1990). Effects of human growth hormone in men over 60 years old. *New England Journal of Medicine*, *323*, 1–6.

- Salmon, A. B., Richardson, A., & Pérez, V. I. (2010). Update on the oxidative stress theory of aging: Does oxidative stress play a role in aging or healthy aging? *Free Radical Biology & Medicine*, *48*, 642–655.
- Schächter, F., Faure-Delanef, L., Guénot, F., Rouger, H., Froguel, P., Lesueur-Ginot, L., & Cohen, D. (1994). Genetic associations with human longevity at the APOE and ACE loci. *Nature Genetics*, *6*, 29–32.
- Schernhammer, E. S., Holly, J. M., Pollak, M. N., & Hankinson, S. E. (2005). Circulating levels of insulin-like growth factors, their binding proteins, and breast cancer risk. *Cancer Epidemiology, Biomarkers and Prevention*, *14*, 699–704.
- Shumaker, S. A., Legault, C., Rapp, S. R., Thal, L., Wallace, R. B., Ockene, J. K., ... WHIMS Investigators. (2003). Estrogen plus progestin and the incidence of dementia and mild cognitive impairment in postmenopausal women: The Women's Health Initiative Memory Study: A randomized controlled trial. *Journal of the American Medical Association*, *289*, 2651–2662.
- Smith, D. W. E. (1996). Cancer mortality at very old ages. *Cancer*, *77*, 1367–1372.
- Sohal, R. S., Sohal, B. H., & Brunk, U. T. (1990). Relationship between antioxidant defenses and longevity in different mammalian species. *Mechanisms of Ageing and Development*, *53*, 217–227.
- Song, Z., von Figura, G., Liu, Y., Kraus, J. M., Torrice, C., Dillon, P., ... Lenhard Rudolph, K. (2010). Lifestyle impacts on the aging-associated expression of biomarkers of DNA damage and telomere dysfunction in human blood. *Ageing Cell*, *9*, 607–615.
- Sprott, R. L. (2011). Introduction: Animal models of aging: Something old, something new. *The ILAR Journal*, *52*, 1–3.
- Stanta, G., Campagner, L., Cavallieri, F., & Giarelli, L. (1997). Cancer of the oldest old. What we have learned from autopsy studies. *Clinics Geriatric Medicine*, *13*, 55–68.
- Strong, R., Miller, R. A., Astle, C. M., Floyd, R. A., Flurkey, K., Hensley, K. L., ... Harrison, D. E. (2008). Nordihydroguaiaretic acid and aspirin increase life span of genetically heterogeneous male mice. *Ageing Cell*, *7*, 641–650.
- Suh, Y., Atzmon, G., Cho, M.-O., Hwang, D., Liu, B., Leahy, D. J., ... Cohen, P. (2008). Functionally significant insulin-like growth factor I receptor mutations in centenarians. *Proceedings of the National Academy of Sciences of the United States of America*, *105*, 3438–3442.
- Tang, M. X., Jacobs, D., Stern, Y., Marder, K., Schofield, P., Gurland, B., ... Mayeux, R. (1996). Effect of oestrogen during menopause on risk and age at onset of Alzheimer's disease. *Lancet*, *348*, 429–432.
- Terry, D. F., Sebastiani, P., Anderson, S. L., & Perls, T. T. (2008). Disentangling the roles of disability and morbidity in survival to exceptional old age. *Archives of Internal Medicine*, *168*, 277–288.
- Thinggaard, M., Jacobsen, R., Jeune, B., Martinussen, T., & Christensen, K. (2010). Is the relationship between BMI and mortality increasingly U-shaped with advancing age? A 10-year follow-up of persons aged 70–95 years. *Journal of Gerontology. Series A: Biological Sciences and Medical Sciences*, *65*, 526–531.
- van Vliet, P., van de Water, W., de Craen, A. J. M., & Westendorp, R. G. J. (2009). The influence of age on the association between cholesterol and cognitive function. *Experimental Gerontology*, *44*, 112–122.

- Varady, K. A., & Hellerstein, M. K. (2007). Alternate-day fasting and chronic disease prevention: A review of human and animal trials. *American Journal of Clinical Nutrition*, *86*, 7–13.
- Waters, D. J. (2011). Aging research 2011: Exploring the pet dog paradigm. *The ILAR Journal*, *52*, 97–105.
- Waters, D. J., Bostwick, D. G., & Murphy, G. P. (1998). Conference summary: First International Workshop on Animal Models of Prostate Cancer. *Prostate*, *36*, 47–48.
- Waters, D. J., Kengeri, S. S., Clever, B., Booth, J. A., Maras, A. H., Schlittler, D. L., & Hayek, M. G. (2009). Exploring mechanisms of sex differences in longevity: Lifetime ovary exposure and exceptional longevity in dogs. *Aging Cell*, *8*, 752–755.
- Waters, D. J., & Wildasin, K. S. (2006) Cancer clues from pet dogs. *Scientific American*, *295*, 94–101.
- Weaver, I. C. G., Cervoni, N., Champagne, F. A., D'Alessio, A. C., Sharma, S., Seckl, J. R., ... Meaney, M. (2004). Epigenetic programming by maternal behavior. *Nature Neuroscience*, *7*, 847–854.
- Weaver, I. C. G., Meaney, M. J., & Szyf, M. (2006). Maternal care effects on the hippocampal transcriptome and anxiety-mediated behaviors in the offspring that are reversible in adulthood. *Proceedings of the National Academy of Sciences of the United States of America*, *103*, 3480–3485.
- Williams, G. (1957). Pleiotropy, natural selection, and the evolution of senescence. *Evolution*, *11*, 398–411.
- Wong, E., & Cuervo, A.M. (2010). Integration of clearance mechanisms: The proteasome and autophagy. *Cold Spring Harbor Perspectives in Biology*, *2*, a006734.
- Wu, D., Cypser, J. R., Yashin, A. I., & Johnson, T. E. (2008). The U-shaped response of initial mortality in *Caenorhabditis elegans* to mild heat shock: Does it explain recent trends in human mortality? *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *63*, 660–668.
- Wu, L. L., Chiou, C. C., Chang, P. Y., & Wu, J. T. (2004). Urinary 8-OHdG: A marker of oxidative stress to DNA and a risk factor for cancer, atherosclerosis and diabetics. *Clinical Chimica Acta*, *339*, 1–9.
- Zuliani, G., Cavalieri, M., Galvani, M., Volpato, S., Cherubini, A., Bandinelli, S., ... Ferrucci, L. (2010). Relationship between low levels of high-density lipoprotein cholesterol and dementia in the elderly. The InChianti study. *Journal of Gerontology Series A: Biological Sciences and Medical Sciences*, *65*, 559–564.